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Chromosomal *ampR* Regulates Plasmid-Mediated Antibiotic Resistance and Gene Duplication Amplification in *Enterobacter cloacae*

Content

Background and Objectives: Gene duplication and amplification (GDA) is crucial for bacterial adaptation to antibiotic pressure [1]. In *Enterobacter cloacae* strain IMT49658-1, ceftazidime induces GDA of a genomic region containing *blaDHA* and adjacent *ampR*(DHA) [2]. The strain also carries plasmid-borne *blaTEM* and *blaCTX*, and chromosomal *blaACT* with *ampR*(ACT). Given the regulatory role of *ampR* in β -lactamase expression, we aimed to investigate whether *ampR*(ACT) influences GDA and the expression of plasmid-borne resistance genes, thereby modulating antibiotic susceptibility and bacterial fitness.

Methods: *ampR*(ACT) was deleted via λ Red recombineering, confirmed by PCR and sequencing. Antibiotic susceptibility was tested by agar disc diffusion assay. GDA copy number was quantified by qPCR of genomic. Meanwhile, Expression of *blaDHA* and *ampR*(DHA) was measured by RT-qPCR. ScanLag was used to assess colony appearance and growth times.

Results: Deleting *ampR*(ACT) unexpectedly increased ceftazidime resistance, despite reduced GDA. RT-qPCR showed *blaDHA* upregulation and *ampR*(DHA) downregulation, suggesting altered regulation. ScanLag revealed delayed growth, indicating a fitness cost.

Conclusions: These findings indicate that *ampR*(ACT) functions as a key regulatory element, influencing both chromosomal and plasmid-borne resistance genes. This study provides new insights into the genetic and regulatory mechanisms shaping antibiotic resistance evolution in *E. cloacae*.

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